CHROMOSOMAL DISEASES IN THE HUMAN PATHOLOGY (review)

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Background. Chromosomal diseases are the cause of 45-50 % of multiple birth defects. Basic research on mutations is performed using genomic technologies to identify a correlation between genotype and phenotype in aneuploidies and to understand its pathogenesis.

Objective. The aim of the research is to study the etiology, pathogenesis of symptoms and diagnostics for patients with Down, Klinefelter, Turner syndromes and double aneuploidies by 21 and sex chromosomes.

Methods. A literature review by the keywords “Down syndrome”, “Klinefelter syndrome”, “Turner syndrome”, “double aneuploidy” for the period of 2000-2020 was carried out.

Results. Down, Klinefelter and Turner syndromes are the most common aneuploidy among viable newborns. Frequency of meiotic non-disjunction events causing these aneuploidies increases with the age of a woman. Identified genes are responsible for pathogenesis of symptoms in trisomy 21, Turner and Klinefelter syndromes. Diagnostics of chromosomal diseases includes prenatal screening programs and postnatal testing.

Conclusions. Cytogenetic variants of Down syndrome are simple complete trisomy 21, translocation form and mosaicism. Trisomy 21 is associated with advanced maternal age. Phenotypic manifestations of Down syndrome are associated with the locus 21q22. The maternal and parental nondisjunction of X-chromosomes in meiosis causes Klinefelter and Turner syndromes. These chromosomal diseases are variants of intersexualism with intermediate chromosomal sex. Down-Klinefelter and Down-Turner syndromes are double aneuploidies. Patients have a Down syndrome phenotype at birth, and signs of Klinefelter and Turner syndromes occur during puberty. Diagnosis of aneuploidy is based on the cytogenetic investigation (karyotyping), DNA analysis, ultrasonography and biochemical markers of chromosomal pathology.

KEY WORDS: Down syndrome; Klinefelter syndrome; Turner syndrome; double aneuploidy.

Introduction

Changes in the number and structure of chromosomes take place in 1% of newborns. They are the cause of 45-50% of multiple birth defects, about 36% of cases of mental retardation, 50% of primary amenorrhea in women, and 10% of sterility in men [1]. Among all cases of birth defects in the life of stillbirths in Ukraine for 2002-2015, almost every 2nd case referred to chromosomal pathology. The main clinical chromosomal syndromes include: Down syndrome (DS) – trisomy 21 – prevalence 1:400-1:1,500 newborns, Klinefelter syndrome (KS) – karyotype 47, XXY with a frequency of 1:500-1,000 newborn boys, Turner syndrome (TS) – karyotype 45,X with a frequency of 1:2,000-5,000 newborn girls [2-6]. Among the viable newborns, 1 of 400 boys and 1 of 650 girls have different forms of aneuploidy by sex chromosomes. KS and TS are most common among them [7].

Double aneuploidy is a rare chromosomal abnormality detected after birth since most reported cases are presented in the form of one lethal aneuploidy and end in early miscarriage. It is extremely rare to find living newborn cases with double autosomal trisomy or autosomal trisomy with sex chromosome monosomy or trisomy [8]. The most frequent co-occurrence is that of DS and KS (coincidence 0.098%), because they are common and relatively well tolerated chromosome abnormalities in humans [9].

The frequency of mutations that causes chromosomal diseases does not depend on race or country. The dynamics of the spread of chromosomal diseases over the past two decades in the world has changed due to the discovery of new cases of double aneuploidy in viable newborns. Early prenatal diagnosis of trisomy 21 as well as postnatal aneuploidy by sex chromosomes is urgent.

Basic research on aneuploidy is conducted using genomic technologies to identify a correlation between genotype and phenotype in chromosomal diseases and to understand its pathogenesis. For diagnosis of hereditary dise-
ases, the medical genetics methods are used, in particular cytogenetic, genealogical, dermatoglyphics, biochemical, and DNA molecular analysis. The cytogenetic method (karyotyping) is widely used for diagnosis of human chromosomal diseases. It is important to be aware of diagnostic possibilities of prenatal testing for early detection of fetal chromosomal pathology such as screening programs for women at early pregnancy. The more deeply the nature of human heredity is analyzed, the more it is realized regarding the methods of diagnostics, treatment and prevention of diseases.

The aim of the research is to study the cytogenetic mechanisms of occurrence, pathogenesis of changes in the phenotype and medical genetic counseling for patients with DS, KS and TS.

**Methods**

A literature review by the keywords “Down syndrome”, “Klinefelter syndrome”, “Turner syndrome”, “double aneuploidy” for the period of 2000-2020 was carried out.

**Review**

According to the literature, in 1866 English physician John Landon Down identified a group of patients with dementia and peculiar somatic abnormalities and called the disease “Mongoloid idiopathy”. The hypothesis of hereditary origin of the syndrome was considered questionable, although in 1932 the ophthalmologist and geneticist Waardenburg suggested that the basis of the pathology may have been “chromosomal insufficiency” [2-3]. Despite this, the cause of this syndrome was clarified only in 1959 by a French scientist Lejeune. He stated that in the karyotype of those patients there were 47 chromosomes, and a small acrocentric chromosome in the group G (21 or 22) was superfluous. As a result of development of the fluorescence chromosome identification method in 1971, it was possible to differentiate 21 of 22 chromosomes and establish the true karyotype in DS. The incidence of patients’ birth with trisomy 21 is not affected by sexual, racial, geographical or population differences. The incidence of children with DS depends on the age of the mother (Table 1) [1]. More than 80% of children with trisomy 21 are born to women under 35 years of age [3]. Hence, prenatal screening and diagnosis of trisomy 21 for a fetus are important for pregnant women aged 35 years and older [10].

Cytogenetic variants of DS are diverse. However, the main part (94-95%) is the cases of simple complete trisomy 21 as a consequence of the nondisjunction of chromosomes to the opposite poles of the cell in the anaphase I or anaphase II of the meiosis. The contribution of maternal nondisjunction is 95 %, and the father’s only 5 %. No dependence on father’s age was found in cases of DS where the additional chromosome originated from father. About 75% of maternal nondisjunction occur during meiosis I, and 25% – during meiosis II, but about 25% of paternal nondisjunction occur during meiosis I, and 75% – during meiosis II [14]. As a result, gametes (oocyte or sperm) have 22 chromosomes (-21) or 24 chromosomes (+21). When a normal gamete that has 23 chromosomes fuses with gametes with 24 (+21) chromosomes, then a zygote 47,+21 is formed, and a child with DS may be born. The karyotype of patients with this will be 47,XX,+21 or 47,XY,+21 [11]. About 2-5% of patients with DS have mosaic forms due to the mitotic nondisjunction of the chromosomes after fertilization (post zygotically), in which one part of the cells has a normal amount of chromosomes (46), and the other is aneuploid (47). The karyotype of patients with 47,XX,+21/46 or 47,XY,+21/46 [11, 13]. The frequency of gonadal mosaicism in trisomy 21, according to some researchers, is actually larger. Available for tissue analysis (blood lymphocytes, skin fibroblasts) have a low mosaic level, while an abnormal line is present in gonads. Study of QRQ polymorphism of chromosomes in 151 families of Down children showed that in 8 families there is gonadal mosaicism in trisomy 21, with all cases of mothers younger than 35 years old. The frequency of carriage of mosaicism in young families was 6.5% (8/123). Approximately 4% of patients with DS have a translocation form. The patients have a diploid chromosome set, but their morphology is disturbed. This form occurs, as a rule, as a result of the Robertson translocation of surplus

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Table 1. The risk of childbirth with Down syndrome in women of different age groups
21 chromosomes between the acrocentric cells (D/21 and G/21). 25% of translocation forms are inherited from native carriers, and 75% of the translocations are formed de novo [14-15]. The translocation form of the DS is not phenotypically different from the classic trisomy 21. The occurrence of this pathology does not depend on the age of parents, while the parents can be phenotypically healthy carriers of translocation (45,-21,t(15q21q)). Consequently, people with a well-balanced translocation have a high risk of having a child with translocation form of DS (1:3). At normal karyotype of parents, the risk of having a baby with trisomy 21 corresponds to a simple trisomy. If a patient has a translocation variant of DS, then the parent’s karyotype must be explored. In young parents, who have children with DS, balanced chromosomal aberrations (1.95 %) are found in 8 times more often than in the other population. When translocation type 13/21 is revealed in the father, the risk of birth of a sick child is 2.4%, and in the mother – about 10%. If one of the parents has a translocation type 21/21, the risk of having a baby with Down syndrome is 100%. About 10% of mothers of the children with DS have mosaicism. In the case of parents’ mosaicism, genetic risk should be considered about 30% [1]. Other forms of trisomy 21 are 1% of DS. They can be caused by: a) a terminal rearrangement of chromosome 21 around the telomeric region, the final chromosome having two centromeres and satellites on both ends; b) as a component of a double aneuploidy (for example, 48,XYY,+21 or 46,X,+21) [14, 16].

Chromosome 21 is the smallest human chromosome and contains 200 to 300 genes. Analysis of the chromosome revealed 127 established genes, 98 predicted genes and 59 pseudogenes [14]. The most acceptable theory for the pathogenesis of trisomy 21 is the dosage hypothesis, which declares that all changes are due to the presence of an extra copy of chromosome 21 within region 21q22. Molecular studies are aimed at determining the region of 21 chromosomes, which would code for phenotypic manifestations of DS. These gene products include morphogens, cell adhesion molecules, components of multi-subunit proteins, ligands and their receptors, transcription regulators and transporters. In particular, it has been found that the DYRK gene (21q22.13) is responsible for mental retardation. Surplus of this gene in mice causes defects in learning and memory. The third copy of the other APP gene (21q21.2) encoding the β-amyloid synthesis causes Alzheimer’s syndrome in the patients with trisomy 21 of over 40 years of age. If as a result of mutation this gene in the patients with DS is absent, the signs of Alzheimer’s syndrome are not manifested [1, 3]. Gart-gene is also detected in the 21 chromosome which in trisomy causes an increased level of purines in the blood of patients that is one of the causes of mental retardation [17]. By the results of expression of this gene you can control the occurrence of DS even in the embryonic period.

The complex of congenital abnormalities specific for DS causes the clinical picture “all children from one family”. Pregnancy, from which patients are born, often is accompanied by toxicosis, the risk of miscarriage. In sick mothers there is an unfavorable obstetric anamnesis (miscarriages, stillborn). The average length of pregnancy is slightly lower than normal. Children with DS are born with moderate pronounced hypoplasia (mass 8-10% below average values) [18]. Craniofacial dysmorphia is manifested by brachycephaly (81%), flat occiput (78%), flattened face, flat bridge of nose (52%), small short nose (40%), large fissured tongue (50%), open mouth (65%), hypoplasia of the upper jaw, high palate (58%), small lowly placed deformed ears, mongolism, epicanthus (80%). Incorrect growth of the teeth, diastema, transverse striae on the lips are typical. In 68% of patients, there are spots of Brushwild (light spots on the iris), 32.2% – cataract. In many patients with trisomy 21, there is a short neck (45%), newborn neck folds, broad fingers and palms, broad toes (70%) [1]. Approximately 40-50% of the affected children have congenital heart diseases (CHD), one of the major causes of morbidity and mortality [19-20]. In Western European countries and the USA, endocardial cushion defect (43%), which results in ventricular septal defect (VSD) (32%), secundum atrial septal defect (ASD) (10%), tetralogy of Fallot (6%) and isolated patent ductus arteriosus (4%) are present in this group of patients [14]. In Asia, isolated VSD has been reported to be the most common cardiac defect (40%) [21]. A study from Korea showed that ASD was the most common defect accounting for 30.5 % of DS, followed by VSD (19.3%), patent ductus arteriosus (17.5%) and atrioventricular septal defect (9.4%) [22]. The second type of ASD was the most common cardiac lesion in Latin America [23-24]. In Libya, the most common isolated cardiac lesion was the ASD, found in 23% of the patients [25]. Molecular studies of rare individuals with CHD and partial duplications of chromosome
21 established candidate gene DSCAM, which was manifested in the heart during cardiac development [26]. About half of the atrioventricular septal defects AVSDs occur in the patients with DS [27]. In the patients with DS and complete AVSD potentially damaging variants in six genes: COL6A1, COL6A2, CRELD1, FBLN, FRZB, GATA5 were identified [28].

Immune problems are also typical in the form of immunodeficiency. Laboratory abnormalities can be revealed in both humoral and cellular immunity. DS is manifested by high incidence of leukemia. However, the children with DS have a 20-fold greater risk for acute lymphoid leukemia and a 50-fold higher risk for myeloid leukemia compare to the children without DS [29]. Development of DS brain is associated with reduction in the neuronal number and abnormal neuronal differentiation. It has been previously reported that DS neuron degenerate subsequently and undergoes apoptosis [4]. The mental retardation with full trisomy 21 is found in almost all patients, and it is basically oligophrenia at the stage of imbecility (65-90%). The average IQ of 10 years old and over is 24. In a mosaic variant of DS, moderate mental retardation is much more common, and normal intelligence is also possible. In the United States DS is 10% of all cases of mental retardation. DS is associated with epilepsy in 10% of cases. Hearing loss and anomalies of the vision organs is often observed in child patients, in particular, cataracts in 66% of the children over 8 years of age [1]. The patients with DS gain more weight compare to other population, and most adults with DS are overweight. Antithyroid antibodies predispose to disturbance of gametogenesis and non-disjunction of the 21 pair of chromosomes. The hypofunction of the thyroid gland in parents is a risk factor. Nowadays, cardiac surgery, vaccinations, antibiotics, thyroid hormones, leukemia therapies, and anticonvulsive drugs (e.g., vigabatrin) have significantly improved the quality of life of individuals with DS. Actually, life expectancy that was hardly 30 years in the 1960s is now increasing – more than 60 years of age [11, 30]. Oral care by dentists for the children with DS from the first months of life involve palatal plates that aim at activating and stimulating different functions, mainly for the orofacial motor function and expression, including language. This is complementary for that of speech therapists, physiotherapists, psycho-motor therapists, and pediatricians [31-32].

Reproductive capacity in DS is described in women, with 40% being ovulatory disorders. Men with this syndrome are sterile. Women with DS produce half normal eggs and half with 24 chromosomes. Consequently, the probability of zygote with trisomy 21 in women with DS is 50%. As about 75% of these pregnancies are completed with spontaneous abortion, the risk to birth of a sick child is less than 50%. Trisomy 21 among aborted embryos is found at a frequency of 1:40. The cytogenetic examination in children with DS is indicated with suspicious of trisomy 21 and with clinically established diagnosis, as the patient’s cytogenetic characteristics from parents and their relatives are needed to predict the health of future children [1].

Diagnostics of DS includes prenatal and postnatal testing. For pregnancies, the high risk of DS is evaluated by fetal sample analysis after invasive chorionic villus sampling (CVS) and amniocentesis, and by laboratory techniques such as conventional karyotype analysis, Fluorescence in Situ Hybridization, Quantitative Fluorescence-Polymerase Chain Reaction, Multiplex Ligation Probe Assay and Array Comparative Genomic Hybridization, which are common techniques used for prenatal diagnosis of DS and each of them presenting with advantages and disadvantages. There is also a noninvasive technique for detection of trisomy 21 by analysis of extracted cell-free fetal DNA screening from maternal plasma samples [14]. The most common screening method contains the measurement of a combination of factors: advanced maternal age, multiple second trimester serum markers and second trimester ultrasonography. The first reported marker associated with DS was the thickening of the neck area. 40-50% of the affected fetuses have a thickened nuchal fold measuring ≥6 mm in the second trimester. When screening by nuchal translucency (NT), about 83% of trisomy 21 pregnancies were identified in the first trimester. Later, it was revealed that screening by a combination of maternal age, NT and biopsy (PAPP-A with second trimester β-hCG) or tri-test [alpha-fetoprotein (AFP), estriol and free β-hCG] has a potential sensitivity of 94% for a 5% false positive rate [4, 33]. However, Ruei-Yu Lan et al, reported about multicenter study of 29,137 cases that enrolled the chromosomal abnormality screening from Taipei city. The mean maternal age of screen-positive group was 34±4.2 years old. The first trimester had 891 cases screening positive with a detection rate of 97.5% for fetal trisomy 21 and false
positive rate of 3.5%. Second-trimester had 334 cases screening positive, the detection rate and false positive rate were 33.3% and 6.4% respectively for trisomy 21 [10].

More recent advances in genomics and related technologies have resulted in development of a noninvasive prenatal screening (NIPS) test using cell-free fetal DNA sequences isolated from a maternal blood sample. Almost 5-10% of DNA in maternal serum are of fetal origin. Fetal trisomy detection by DNA-analysis from maternal blood has been done using massively parallel shotgun sequencing (MPSS). By next generation sequencing platforms, millions of amplified genetic fragments can be sequenced in parallel. MPSS detects higher relative amounts of DNA in maternal plasma from the fetal trisomy chromosome compare with reference chromosomes. Platforms differ according to whether amplified regions throughout the genome, chromosome-specific regions, or single nucleotide polymorphisms (SNPs) are the targets for sequencing [34]. Although studies are hopeful and exhibit high sensitivity and specificity with low false-positive rates, there are drawbacks to NIPS. Specificity and sensitivity are not consistent for all chromosomes; this is due to different content of cytosine and guanine nucleotide pairs. False-positive screening results take place because the sequences derived from NIPS are derived from the placenta, like in CVS they may not reflect the true fetal karyotype. Therefore, currently invasive testing is recommended for confirmation of a positive screening test and should remain as option for patients seeking a definitive diagnosis. NIPS began as a screen for only trisomy 21 (T21) and rapidly developed to include other common aneuploidies for chromosomes 13 (T13), 18 (T18), X, and Y [35, 36]. Amniocentesis is the most conventional invasive prenatal diagnostic method accepted in the world. It is mostly performed to acquire amniotic fluid for karyotyping from 15 weeks onwards. Actually, amniocentesis performed before 15 weeks of pregnancy is referred to as early amniocentesis. CVS is usually performed between 11 and 13 weeks of gestation and includes aspiration or biopsy of placental villi. Amniocentesis and CVS are quite reliable but increase the risk of miscarriage up to 0.5 to 1% compare with the background risk [33].

A number of European countries have established a network of OSCAR clinics (One Step Clinic for Evaluation of Risk for Fetal Abnormalities). Clinics of “one visit” allow calculating the individual risk of a child’s birth with chromosomal pathology regarding of the age of pregnancy, ultrasound and biochemical markers of chromosomal pathology. In particular, among the biochemical markers in the first trimester, the content of plasma protein-associated protein (PAPP-A) and β-chorionic gonadotropin (β-hCG) levels are determined [18]. For non-invasive prenatal diagnosis, studies of fetal trophoblastic cells from the cervical canal of women in the first trimester of pregnancy are also used [37]. Constantly conventional karyotype from peripheral blood in postnatal period is performed to confirm diagnosis for all patients suspected with DS [14].

Polysomy by sex chromosomes has less significant pathological manifestations than aneuploidy by autosomes. It may be explained by the fact that the Y chromosome contains little genes, and additional X chromosomes may be heterochromatic. Genetic intersexuality is the result of sex chromosome aneuploidies. TS and KS are variants of intersexuality with an intermediate sex [7].

TS (45,X) is a single form of monosomy in the liveborns. Monosomy X was discovered by Henry Turner in 1938 (earlier, in 1930, the description was made by Otto Ullrich). Cytogenetic syndrome 45,X was discovered by SE Ford in 1959 [5]. Clinically TS manifests itself in three directions: 1) hypogonadism, underdeveloped genital organs and secondary sexual characteristics; 2) congenital defects; 3) short stature. TS is one of the clinical forms of sexual differentiation disorders – agenesis with somatic malformations. In the reproductive system there is a lack of sexual glands (agenesis of gonads), hypoplasia of the uterus and fallopian tubes, primary amenorrhea, inadequate hair follicles in the pubic and axillary areas, underdevelopment of the mammary glands, deficiency of estrogens, and excess of pituitary gonadotropins. In children with TS, congenital malformations of the heart, vessels and the kidneys make up to 25%. In newborns, the neck is short with excess skin and wing-like folds, with low neck hair growth. These patients have lymphatic edema of the feet, legs, hands and forearms. At school age, especially at the puberty period, there is a lag in the growth and development of secondary sexual characteristics. In 16-50% of cases the patients with TS are mentally retarded, prone to the lymphedema and Hashimoto’s thyroiditis. It is established that if a TS patient received an X-chromosome from their father, they would have a higher IQ and better adaptation in so-
ciety than that received from the mother’s sex chromosome. Chromosomal anomalies in women with TS are quite different: 50% of these patients have 45,X karyotype in blood lymphocytes, 30-40% are mosaics in most cases with a karyotype 45,X/46,XX, less than 45,X/46,XY; 45,X/47,XXX. Among liveborn limited placental mosaicism is often observed. The more normal cells a fetus has the greater chance of its survival. About 10-20% of patients with TS have structural changes in the X-chromosome, which include deletion of short or long arms [46,Xp-; 46,Xq-], isochromosomes [46,X,Xq]; 46,X,i(Xp)], ring chromosomes [46,X,R(X)].

These varieties of chromosomal aberrations stipulate for the phenotype of patients. Molecular studies have shown that 60-80% of monosomies X are due to the disappearance of the paternal sex chromosome that occur either in early mitosis during embryogenesis or in the process of meiosis. A repeated risk of birth of a sick child with TS in a family with normal parents karyotype does not exceed 1% [7]. Molecular analysis also allowed precise determination of specific genes responsible for the phenotype of TS. For example, the mutation in the SHOX gene causes low growth. This gene is localized in the distal segment of the X chromosome (Xp22.33) and in the short arm of the Y chromosome. Consequently, this gene undergoes normal transcription being in two copies of a man and a woman. So, as the patients with TS have only one copy of this gene, they have a high probability of short stature development. According to Linglart et al. [38], hormone reposition in the patients with TS may reduce likelihood of short stature development. Webbed neck, lymphedema or coarctation of the aorta in infancy should prompt a peripheral blood karyotype to rule out TS [39]. The standard 30-cell karyotype is recommended and can detect 10% mosaicism with 95% confidence [5]. Multiple sequences adjacent to the Y-centromere should be amplified using polymerase chain reaction techniques to detect cryptic Y-material. When Y-chromosome material is present in the standard karyotype or on such additional testing (incidence of 5-12%), prophylactic gonadectomy is still recommended by expert consensus, albeit at a lower quality of evidence, due to an increased risk (around 10%) of gonadoblastoma [39-40]. In TS fetal ovaries develop normally in the early stages of embryogenesis, but the absence of a second X-chromosome leads to an accelerated loss of oocytes within 2 years. Spontaneous puberty has been reported in 14% of TS patients with monosomy X and up to in one third of patients with mosaicism [5]. Young TS women with normal ovarian function should be counseled about fertility preservation options. Gonadotropins (especially follicle-stimulating hormone) should be monitored annually starting at about 11 years of age to confirm hypergonadotrophic hypogonadism prior to pubertal induction [6]. Anti-Müllerian hormone (AMH) and inhibin B measurements have also been shown to predict ovarian insufficiency when found to be low, and AMH is perhaps the best indicator of ovarian reserve [41]. However, the majority of the girls with TS require induction of puberty and estrogen/progestin replacement therapy to achieve adequate breast development, uterine maturation and peak bone mass. Transdermal 17-β estradiol (TDE) is now the preferred treatment starting at age around 11-12 years old. Compared with oral estrogens, TDE is thought to be more physiologically delivery since it will avoid the first-pass effect in the liver with improved bioavailability. A recent meta-analysis showed improved whole-body bone mineral density, fasting glucose and total cholesterol with TDE therapy compare with oral estrogens [42].

KS is the most extensive major anomaly of sexual differentiation in men. The disease is described in 1942 by Harry Klinefelter in males with sexual retardation. In 1959, Jacobs and Strang showed the presence in these patients of a pathological karyotype. The most common cause of KS is nondisjunction of XX homologues for meiosis. Meiotic non-disjunction events causing KS are inherited maternally in 50-56% of cases (36% occur in 1st and 20% in 2nd meiosis), and paternally in 44-50% (100% 1st meiosis) [43, 44]. With the age of the mother, as with DS, the probability of birth of a baby with this syndrome increases [1]. In 40-year-old mothers, ill children are born by 2-3 times more often than in 30-year-olds. Preliminary, it is established that half of cases with a karyotype 47, XXY end with spontaneous abortions. Mosaic among liveborn is observed in 15% of cases [7]. The presence of the Y chromosome determines the formation of a male phenotype. Genetic imbalance due to excess X-chromosome is manifested clinically during puberty as underdevelopment of the testicles and secondary male sexual characteristics. The presence of an extra chromosome can cause higher levels of gene expression and gene products in amounts affected by regulation at different levels, pro-
tein degradation and modification [45]. Also, the interaction pattern of the proteins encoded by the additional chromosome can influence the severity of a trisomy. In the case of KS, X chromosome inactivation (XCI) counterweights the extra amount of X chromosome genes (about 2000). Still, roughly 15 % of X-linked genes escape XCI, including genes in the two pseudoautosomal regions (PARs). A total of 22 X chromosome genes were deregulated in KS compared to the controls: 16 were upregulated (AKAP17A, ASMTL, CSF2RA, EIF1AX, EIF2S3, GPR82, GTPBP6, IL3RA, PLCXD1, PPP2R3B, PRKX, RP11-EPO60G15.3, SEPT6, SLC25A6, TMSB4X and XIST) and six were down-regulated (BEND2, BEX1, COX7B, FOXO4, NHS and TFE3). Some of these genes were expressed from PAR1: AKAP17A, ASMTL, CSF2RA, GTPBP6, IL3RA and PLCXD1, but no differentially expressed genes were observed encoded from PAR2. XCI compensates extensively for the extra X chromosome gene dosage in KS males and probably increases their chances of survival compare to other aneuploidies [17].

Thus, in the patients with KS have a high stature, eunuchoidism, narrow arms and wide pelvis, subcutaneous fat development by female type, absent hair growth on the face, in the axillary areas and in the area of the pubis, sparse hair, underdeveloped testicles and their endocrine function (production of androgens) is reduced. The prevalence of androgen deficiency in the patients with KS is 79%. The patients in most cases are sterile (due to azoospermia, oligospermia). Among men, who have infertility, more than 10% have an additional X-chromosome. Approximately one in three patients with this syndrome has gynecomastia and an increased risk of breast cancer development. This risk can be reduced by having a mastectomy. Gynecomastia and obesity with fat deposits for the female type are a result of relative hyperestrogenemia [7]. In the KS patients this may lead to an increased incidence of female predominant autoimmune disorders [46].

The system biology approaches together pointed to novel aspects of KS phenotypes including perturbed AK-STAT pathway, dysregulated genes important for disturbed immune system (IL4), energy balance (POMC and LEP) and erythropoietin signaling in KS. Sex chromatin is detected in 80% of the patients with KS. Modified laboratory parameters: decreased testosterone in the blood and urinary excretion of 17-ketosteroids; an increase in the amount of estrogen in the plasma; as well as their excretion with urine; high levels of gonadotropins in plasma, especially follitropin, are observed. Biopsy of testicles should be used due to the inaccuracy of laboratory parameters (hyalinosis of the walls of seminal tubules, aplasia of the generative epithelium, aspermatia). Usually KS males present a normal growth hormone – insulin-like growth factor IGF-I axis and increased luteinizing hormone and follicle-stimulating hormone levels due to their androgen deficiency [17]. However, early diagnosis of sex chromosome syndromes may provide better development and quality of life for patients with these disorders. Samango-Sprouse et al. [47] observed significant advantages in the cognition, language, intellectual, and psychomotor functions of the patients with KS treated with hormone therapy at 3-6 years of age compare to a group of untreated patients.

The double aneuploidy 48,XXY,+21 was first described in 1959, and the incidence is reported to be 0.4 to 0.9 per 10,000 male births [9, 47, 48]. Most commonly, double aneuploidy occurs when two meiotic nondisjunction events happen from the same or different parental origins. The coincidence rate of DS and KS in the same patient is 0.098 % in newborns, and it has been found to dependent on parental age. Advanced maternal age is one of the main risk factors for double aneuploidy, as it is for single chromosomal aneuploidies caused by parental nondisjunction during gametogenesis [49-51]. The nonrandom aspect of double aneuploidy provides evidence that a hereditary predisposition to nondisjunction exists, with one chromosomal imbalance increasing the risk of another to occur [52]. The phenotype of 48,XXX,+21 double aneuploidy is characterized by classical DS features in children, who then develop additional KS symptoms from the age of puberty [9, 52]. Since the sexual development primarily depends on the presence of the Y chromosome, the 48,XXX,+21 individuals have male phenotype. KS is a form of primary testicular failure with testicular hypotrophy and elevated gonadotropin plasma levels, and it is the most common form of male hypogonadism [53]. In addition, the spectrum of different mental problems could be wider, and their risk could be higher in this group of patients. In cases of combined DS and KS the incidence of congenital heart disease could be higher compare to the isolated forms of these aneuploidies. Sevcan Tug Bozdogan and Atil Bisgin reported of Down-Klinefelter syndrome with accompanying both heart defects and hypothroidism [49].
The combination of DS and TS is very rare (1 in 2,000,000) [15, 54]. Evren Gumus observed a two-year old girl with karyotype from the peripheral lymphocytes using G-bandings “mos45,X[41]/47,XX,+21[59]”. She had stigmata’s such as up slant palpebral fissures, brachycephaly and epicanthus, renal hypoplasia and hepatomegaly. In this case, at early ages, the clinical picture is completely compatible with the autosomal chromosome aneuploidy, but the phenotypic effects of TS will become dominant in post pubertal period [15]. Therefore, Down-Klinefelter and Down-Turner syndromes are rare chromosomal abnormalities. A patient present with physical characteristics of DS at birth but KS or TS features during puberty. It is important to counsel families about recurrence risk for future pregnancies. A clinical geneticist evaluation is also recommended.

Conclusions
Cytogenetic variants of Down syndrome are simple complete trisomy 21, translocation form and mosaicism. Trisomy 21 is associated with advanced maternal age. Phenotypic manifestations of Down syndrome are associated with the locus 21q22. The maternal and parental nondisjunction of X-chromosomes in meiosis causes Klinefelter and Turner syndromes. These chromosomal diseases are variants of intersexualism with intermediate chromosomal sex. Down-Klinefelter and Down-Turner syndromes are double aneuploides. Patients at birth have a Down syndrome phenotype, and signs of Klinefelter and Turner syndromes appear during puberty. Diagnosis of aneuploidy is based on the cytogenetic investigation (karyotyping), DNA analysis, ultrasonography and biochemical markers of chromosomal pathology.

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